

# Expression properties of the novel *Staphylococcus aureus* extracellular protease Jep under infection-relevant stress conditions

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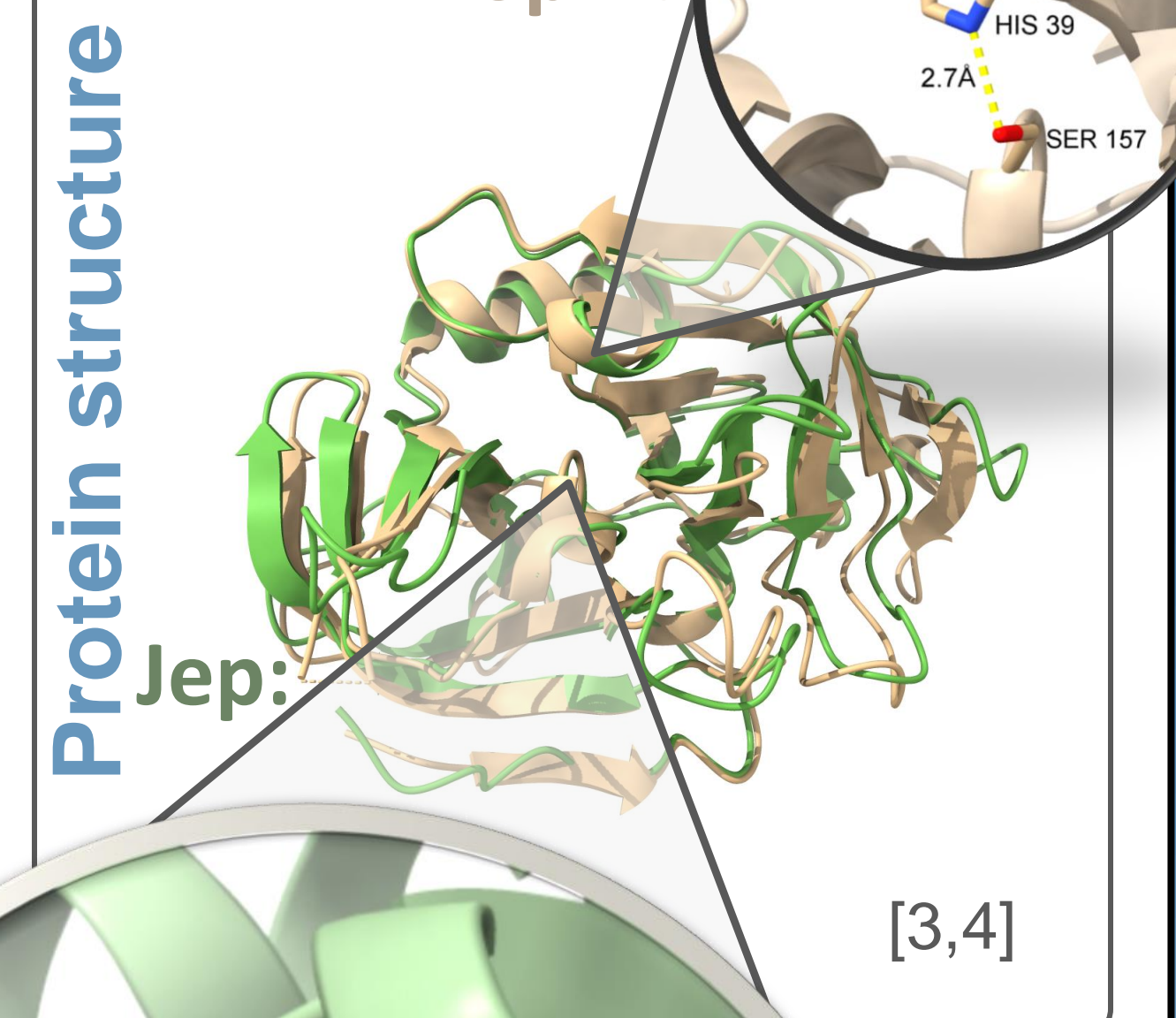
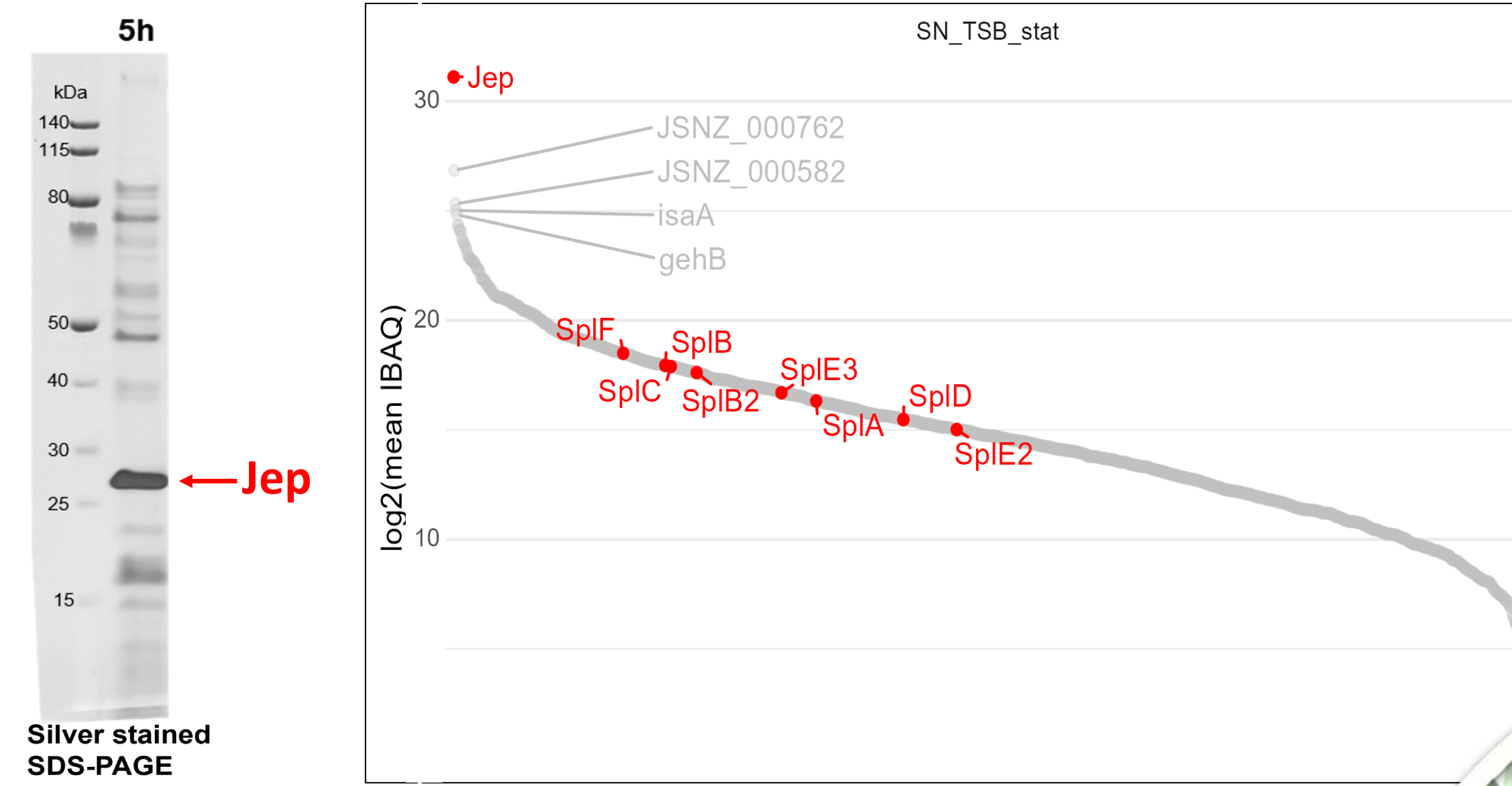
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## A family of proteases

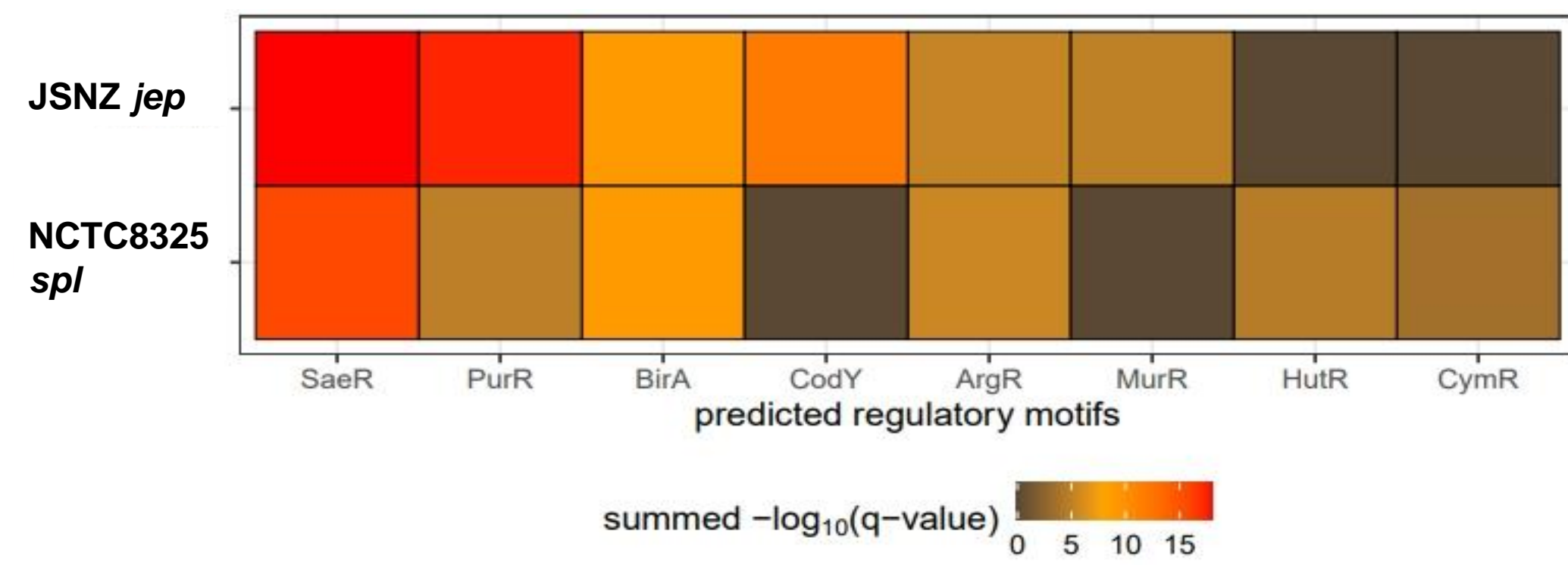
Extracellular proteases are important virulence factors in *Staphylococcus aureus*. This also comprises a set of serine protease-like proteins (Spl) whose role in infection is still poorly understood. In the mouse-adapted *S. aureus* strain JSNZ, we recently identified a closely related protease, JSNZ extracellular protease (Jep) [1,2]. It shares significant sequence homology and a conserved catalytic triad with the Spls, making it an interesting candidate for investigating the role of serine proteases in murine *S. aureus* infection models.

Here, we characterize *jep*-expression in JSNZ under different stress conditions. Furthermore, we optimize a system for inducible *jep*-expression to analyze the effect of the protease in different genetic backgrounds.

## Jep, the dominant protein of the secretome



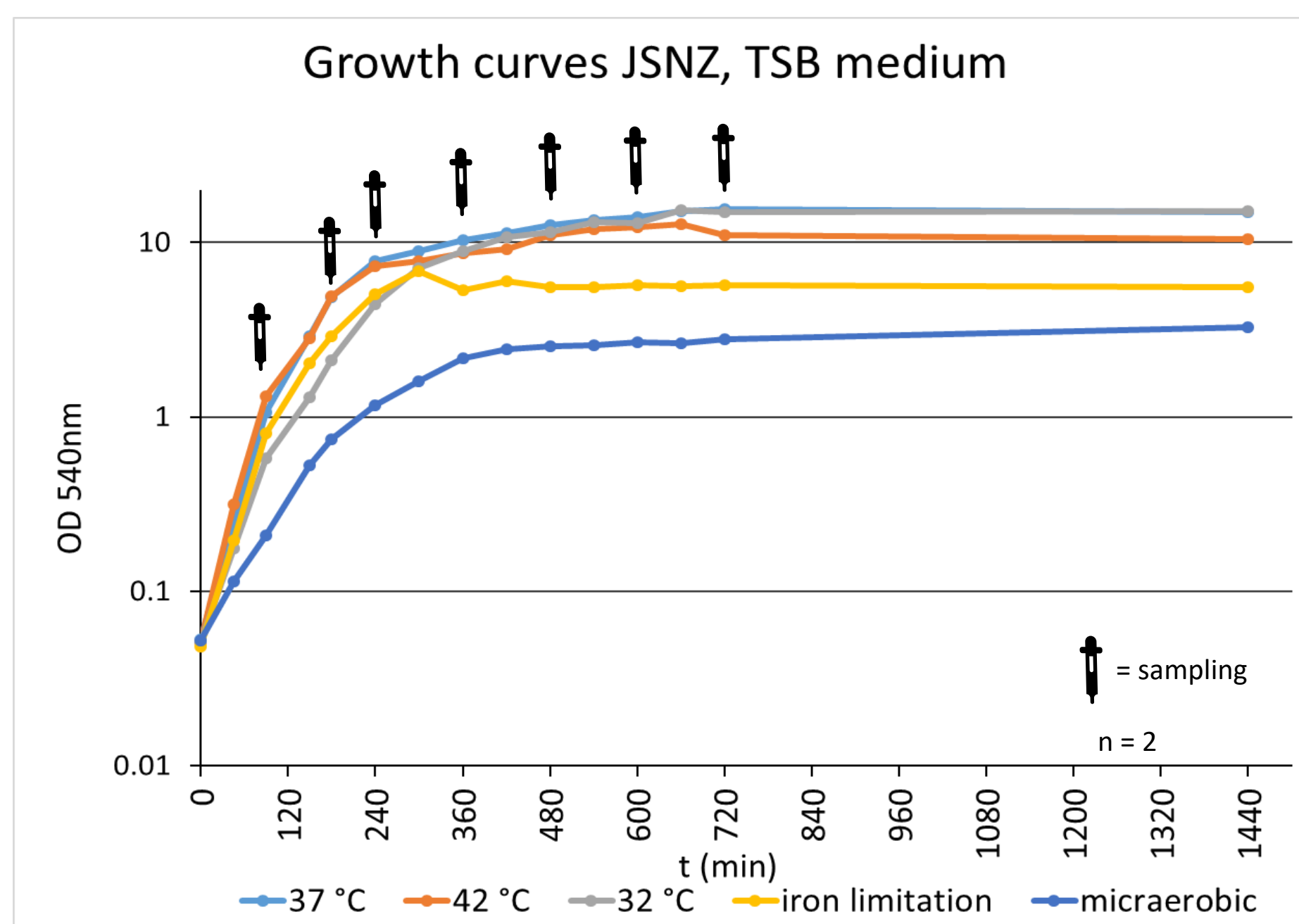
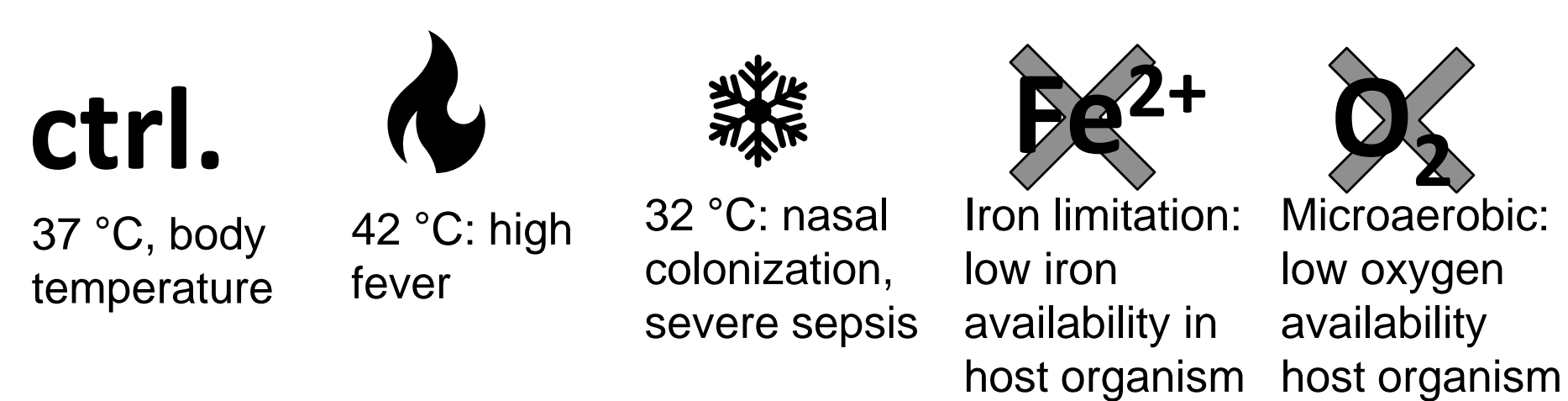
## Regulator prediction



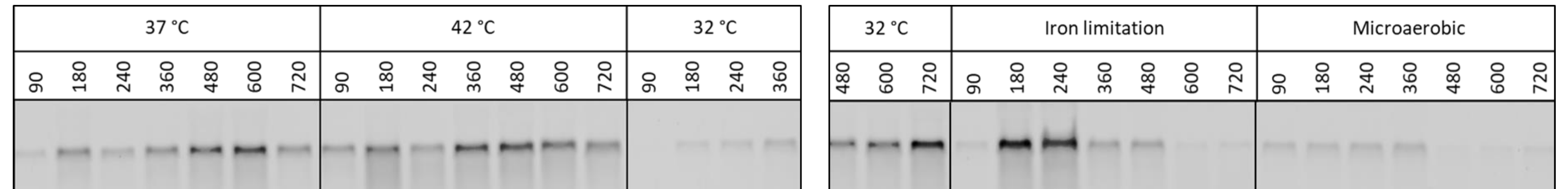
- Analysis performed with *in silico* tool "FIMO" [5], regulatory motifs from "RegPrecise" [6]
- 300 bp upstream – 200 bp downstream of *jep* and *spl* start codon

## Growth phase and stress-dependent *jep* expression

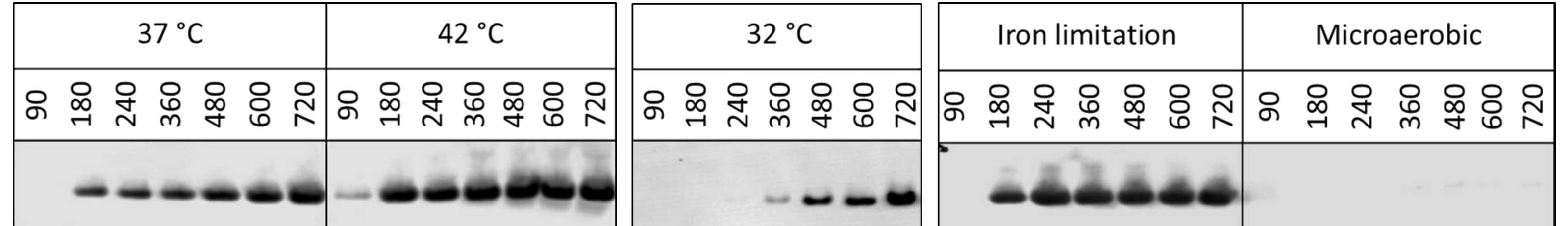
### Infection relevant conditions:



### Northern blot analysis – Mapping gene expression



### Western blot analysis (secretome) – Mapping protein levels

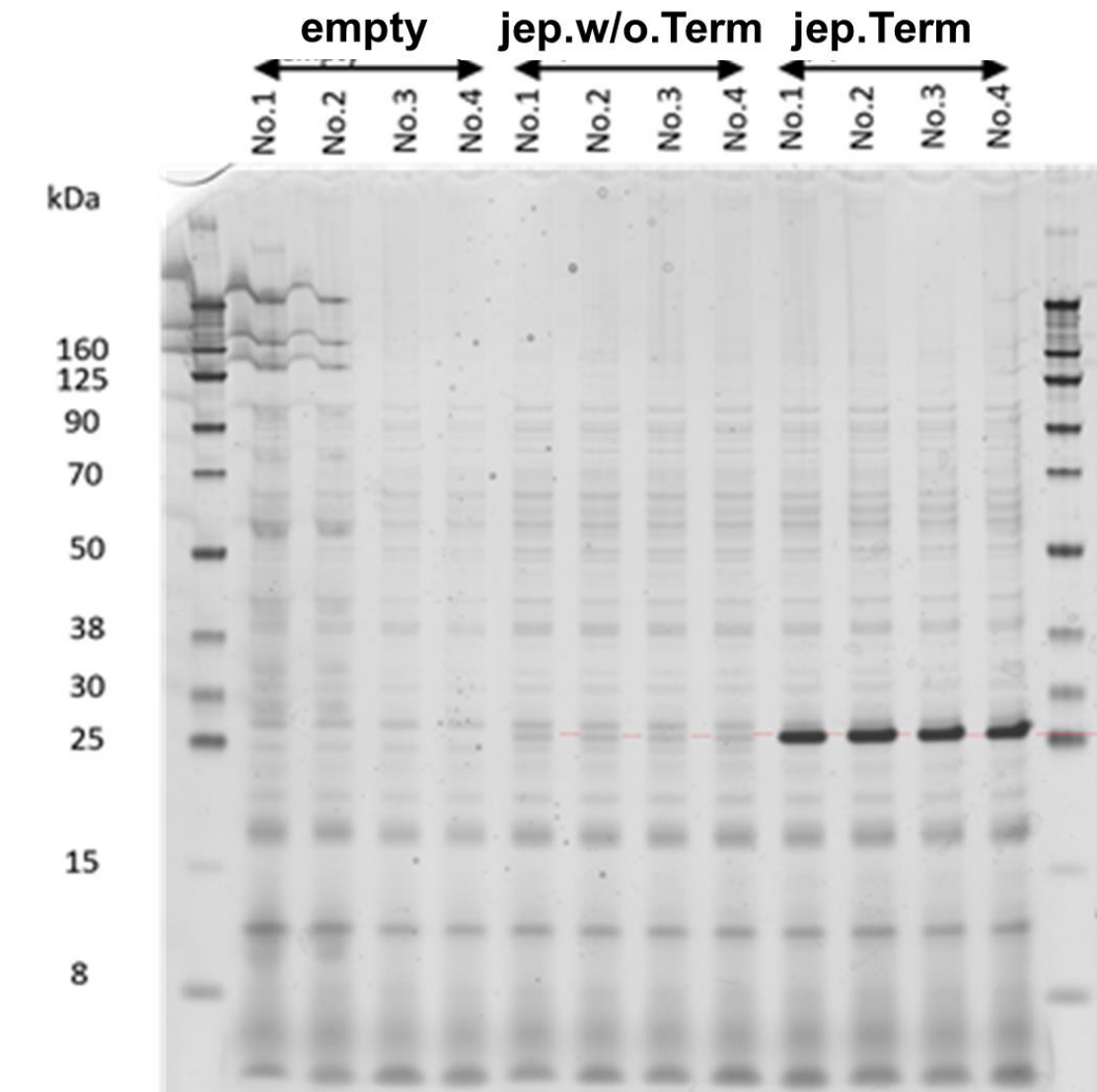
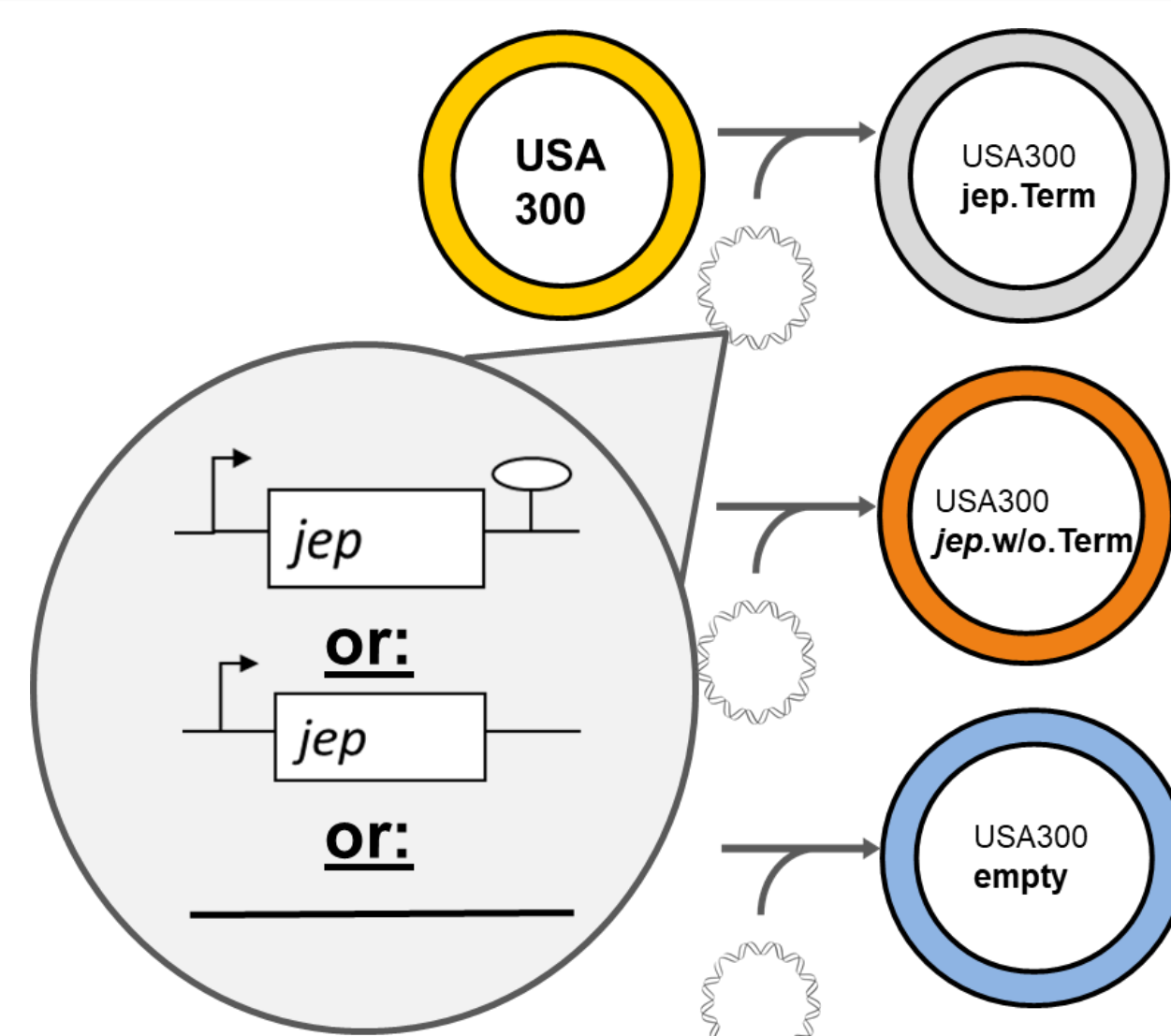
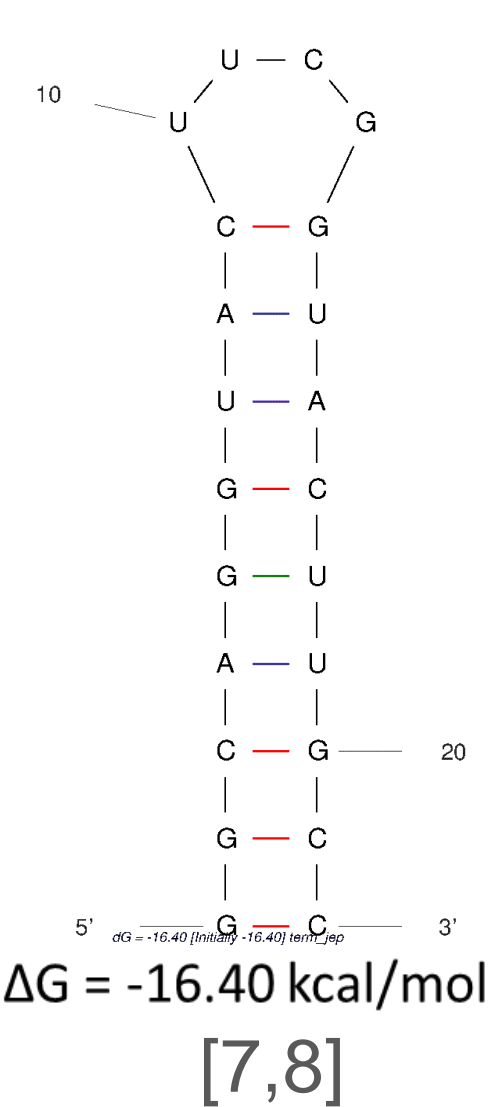


## Results

- Expression of *jep* starts in exponential phase and increases until stationary phase
- The protein accumulates in the supernatant
- Iron limitation and heat stress lead to earlier and higher expression
- Oxygen limitation decreases expression drastically

## Inducing *jep* expression in a human adapted strain

### Identification of potential terminator → Cloning of *jep* sequence with and without terminator → Induced *jep* expression → SDS-PAGE of the supernatant → Northern blot analysis (*jep*)



empty				jep.w/o.Term				jep.Term			
BR1	BR2	BR3	BR4	BR1	BR2	BR3	BR4	BR1	BR2	BR3	BR4

## Summary

- Jep is the most abundant protein in JSNZ stationary phase supernatant indicating a central role in JSNZ lifestyle
- Sequence and structure homology of Jep and Spls: Jep could give insights to the role of Spls in human infection
- Potential regulatory motifs connect *jep*-expression to virulence
- Protein accumulates in supernatant: indicates high stability of the protease
- Iron limitation and heat stress lead to earlier *jep*-expression, oxygen limitation decreases expression: potential adaptation to diverse niche environments in the host organism
- System for inducible *jep*-expression optimized

## Outlook

- Comparison of JSNZ intra- and extracellular proteome under stress conditions shown above via mass spectrometry
- Characterization of *jep*-expression under oxidative stress

## References

1. Holtfreter, Silva et al. "Characterization of a mouse-adapted *Staphylococcus aureus* strain." *PLoS one* vol. 8,9 e71142. 2 Sep. 2013.
2. Schulz, Daniel et al. "Laboratory Mice Are Frequently Colonized with *Staphylococcus aureus* and Mount a Systemic Immune Response-Note of Caution for *In vivo* Infection Experiments." *Frontiers in cellular and infection microbiology* vol. 7 152. 2 May. 2017.
3. Dubin, Grzegorz et al. "Enzymatic activity of the *Staphylococcus aureus* SplB serine protease is induced by substrates containing the sequence Trp-Glu-Leu-Gln." *Journal of molecular biology* vol. 379,2 (2008): 343-56.
4. Knyphausen, Philipp et al. "Evolution of protease activation and specificity via alpha-2-macroglobulin-mediated covalent capture." *Nature communications* vol. 14,1 768. 11 Feb. 2023.
5. Grant, Charles E et al. "FIMO: scanning for occurrences of a given motif." *Bioinformatics (Oxford, England)* vol. 27,7 (2011): 1017-8.
6. Novichkov, Pavel S et al. "RegPrecise: a database of curated genomic inferences of transcriptional regulatory interactions in prokaryotes." *Nucleic acids research* vol. 38 Database issue (2010).
7. Naville, Magali et al. "ARNold: a web tool for the prediction of Rho-independent transcription terminators." *RNA biology* vol. 8,1 (2011): 11-3.
8. Baerends, Richard J S et al. "Genome2D: a visualization tool for the rapid analysis of bacterial transcriptome data." *Genome biology* vol. 5,5 (2004): R37.